

Living up to Life EM Sample Preparation

Techniques and pathways

By Leica Nanotechnology

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SOME TECHNIQUES....

ULTRAMICROTOMY / CRYO-ULTRAMICROTOMY LEICA UC6-FC6	TRIMMING/ KNIFE-MAKER LEICA KMR2	CRITICAL POINT DRYING IN BRAGA?
CONTRASTING /LABELLING	PROCESSING/ DEHIDRATATION CHEMICAL FIXATION RESIN EMBEDING	COATING HIGH VACUUM EVAPORATOR POLARON CRESSINGTON SPUTTER
FREEZE ETCHING / FREEZE FRACTURE	CRYO-TRANSFER	FREEZE SUBSTITUTION
CRYO-FIXATION HIGH PRESSURE FREEZING	CRYO-FIXATION BARE GRID TECHNIQUE PIN INMERSION	CRYO-FIXATION SELF PRESSURE FREEZING
TARGET PREPARTION (CUTTING, MILLING, SAWING, POLISHING, GRINDING) IN MINHO?	ION BEAM CUTTING	ION MILLING SYSTEM









EXAMPLE: Workflow of Room Temperature Specimen Preparation for TEM





EM UC7:Ultramicrotome



EM KMR3 : Knifemaker





The Leica Ultracut EM UC7

- is used for ultrathin sectioning for TEM
- Surface planing of AFM and **SEM samples**. Industrial material manufacturers and research (polymer, rubber, and materials), as well as cosmetic samples.
- Semithin sections for LM and FT-IR
- 3D reconstruction (tomography, serial sectioning)



Leica EM UC7 application





Trimming and /or Targeting



Why using a trimming tool?

Save time and money
Save diamond/glass knives

TRIM2



Synergies - Room temperature SEM workflow

Tissue Processing – Leica EM TP



Automated tissue processing for fixation and dehydration.

Critical Point Drying — Leica EM CPD300



Automated critical point drying for extremely well perserved sample structures.



Liquid CO₂ concentration



Ethanol, Acetone concentration

Coating — Leica EM ACE200 and ACE600

Imaging and analysis



Automated coating for reproducible, thin and conductive layers.





Antenna of male mosquito (Gold sputter coating), SEM.



Material Research Samples



H₂O concentration



Tissue Processing





• WHY ? SAFETY REASONS !!!

TP

1) Automatic processing of tissue:

Minimizes contact with hazardous reagents (eg. Uranyl, Osmiun or Glutaraldehyde during the CHEMICAL FIXATION).

2) Closed processing chamber with fume extraction system



Critical Point Drying



Cells Insects Tissues Wafers Etc...

Why?

Surface Tension of Water Damages Sample, but not CO2 You should avoid water in the EM column



And the result is....





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Courtesy of D. Gruber, University of Vienna, Austria





Mold & amp; Bacteria Laboratories

W. Müller, University of Utrecht, Netherlands

labs.com







Why do we coat ?

SEM

- Reduce charging, inhibits charging
- Improves the secondary electron signal
- Higher electron yield from specimen <u>surface</u>
- Prevent specimen thermal damage

TEM

- Support films for TEM-grids (carbon on formvar)
- Generate contrast on thin samples (low-angle shadowing)
- Freeze-fracture replica technique

Or, to produce conductive layers in Micro-electronics research.



Different Coatings

• WHY METAL SPUTTERING COATING?

- Chemically inert, does not react with specimens or acids
- Stable in the electron beam
- Transparent to the electron beam (does not create contrast!)
- Conductive reduces charging
- WHY CARBON COATING?
- Reduce charging
- Localize secondary electron (SE) and back-scattered electron (BSE) signal to the surface in SEM
- Reduce heating of non-conductive specimens to increase potential exposure time to the electron beam
- Reveal topography with shadowing (TEM or SEM)







Other techniques in your coater:

- **Sputtering metals** like gold, gold/palladium, silver, platinum etc... and additionally to LowVac, HigVac sputtering for materials like Aluminum, Chromium, Iridium, Molybdenum, Titanium Tungsten etc.
- **Conductive carbon films** for X-ray analyses, grids and backing of collodion and formvar films for biological EM via carbon thread /rod evaporation
- Thermal resistance evaporation of metal and carbon.
- **e-beam evaporation**. The finest layers, either carbon or metals.
- **Freeze drying, freeze etching, in combination with a VCT100**: Freeze fracturing **double replica**, cryo-transfer.



Metal coating: Electron beam evaporation

Evaporation of metals using the electron beam gun

- For materials with high melting point
- For very fine grained layers (W, Ta/W, Cr, Pt/C)
- Shadowing
- Homogeneous layers with DARS (Double Axis Rotary Shadowing)
- Low heat transfer to specimen
- No charged particles to your sample







Coating quality depends on:

- Coating technique
- Layer thickness
- Coating material
- Vacuum conditions
- Specimen temperature
- The specimen itself ("decoration effects")
- Effects after coating



Coating quality - Specimen temperature WHY CRYO-COATING ???

- Reorganization of the coating material depends on the surface temperature.
- Colder temperature reduces mobility of the coating material causing:
 - Smaller distance between grains
 - Finer structure



Synergies - Cryo SEM workflow

High Pressure Freezing -Leica EM HPM100, Leica EM PACT2



Superior cryo fixation to observe aqueous biological and industrial samples near to native state.

Leica EM UC7 with EM FC7

Cryo Ultramicrotomy -

Coating -Leica EM ACE600 cryo outfit



Imaging and analysis



High vacuum coating in conjunction Leica EM VCT100 (vacuum cryo transfer) system for the finest metal and carbon layers.

Mouse kidney (Platinum cryosputter coating), Cryo SEM.



High quality ultrathin sectioning/planing for light, electron, and atomic force microscopy examination.











Why cryofixation?

- The cellular constituent is rapidly immobilized
- Cryofixation is a physical fixation of all cellular components simultaneously
- Enzymes and antigens are not denatured (antigenecity)
- Cells retained in their 'native' state
- The physical properties of a frozen sample allow cryosectioning without any additional support by embedding medium (CEMOVIS).



Why High Pressure Freezing?

- No changes in the physical equilibrium because of fast and precise correlation between pressure and temperature
- No cryo-protection needed, so no alteration of the cellular processes
- Vitrification of sample thickness up to 200 μm





- •LN2 at 2100bar used for pressure buildup AND cooling
- •Alcohol is synchronizing pressure buildup and cooling
- •Pressure is the same everywhere

- •Pressure build-up and cooling separated
- •LN2 at 10 bar used for cooling only
- •Pressure transferred only to specimen (2100bar) by filler



Cryo-fixation Plunge Freezing and Metal Mirror



In biological research, virology, protein crystallography, pharmaceutical labs, cosmetic companies and paint research.

Process Automation

- Automatic blotting with optional sensor control
- Controllable ethane container temperature

Reproducibility with high sample quality



Leica EM FC7

- Three different cryo-condition modes:
 - standard mode
 - high gas-flow (to reduce ice contaminations)
 - wet sectioning (for polymer sectioning at wet condition using DMSO), wide range of different temperature settings of knife / specimen and gas (-40°C to -160°C)



SPECIMEN MOUNTING AND TRANSFER Why? How could I transfer my sample?



LOADING STATION









VCT 100 - HIGH VACUUM CRYO TRANSFER TO SEM





Pathway with Freeze Substitution



Dehydration of a cryofixed specimen by exchange of ice with an organic solvent

Freeze substitution is a link between cryofixation and room temperature TEM observation.



Why Freeze Substitution?

• Enables sectioning at room temperature

 Avoids artifacts produced by conventional room temperature procedures

• Preserves antigenicity







Freeze Substitution

- Minimized aggregation and redistribution of diffusible elements
- Fixatives are uniformly distributed throughout the sample



Cryotechniques in Biological Electron Microscopy, 1987, Steinbrecht RA, Zierold K)



FSP

Freeze **S**ubstitution **P**rocessor for EM AFS2

- Automatic reagent handling
- Special embedding moulds for
 - FS of high pressure frozen samples
 - PLT samples

"Load and Leave"





MATERIAL SCIENCES



EM TXP

Target Surfacing Device for SEM, LM, TEM















- For site specific cross sections (target preparation)
- Surfacing of small samples
- Pre-preparation prior to ion beam polishing (SEM)
- Pre-preparation prior to ion beam thinning (TEM)
- Pre-preparation prior to ion beam slope cutting (SEM)
- Pre-preparation priot to ultramicrotomy (SEM, TEM) TRIMMING
- Microelectronic market
- Watch industry
- Material reseach (Nano technoloy)











Ion Milling- Slope Cutting FOR SEM

Triple Ion Beam Cutter TIC3X

Used for SEM microstructure analysis (EDS, WDS, EBSD, CL) and AFM investigations







Triple Ion Source







Principle

- Three ion beams hitting the sample from different directions
- Fixed sample
- Does not substitute a FIB on SEM, just because the area here is huge.
- 1x4mm prepared





Ion Milling – Only for TEM







Applications of the RES 101

TEM

- Plan view preparation
- Cross-sectional sample preparation
- ➢ FIB Cleaning

SEM

- Surface cleaning
- Polishing
- Contrast enhancement
- > Ion beam slope cutting (35° and 90°)

LM



Muito obrigado pela vossa atenção!

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